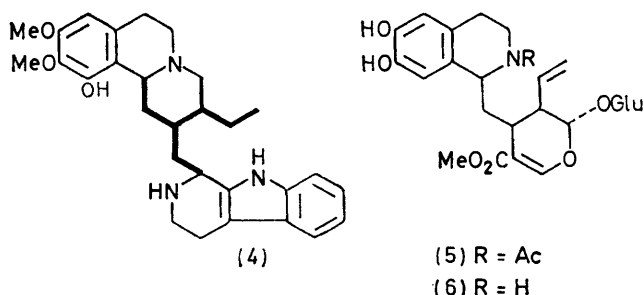
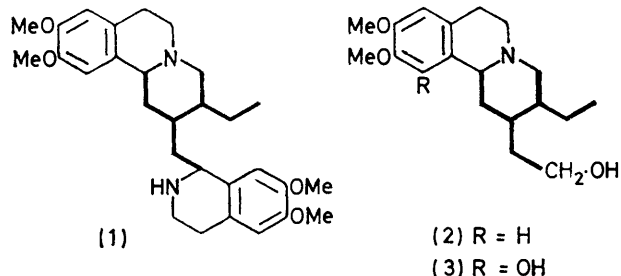


Alangiside, the Monoterpenoid Alkaloidal Glycoside from *Alangium lamarckii* Thw.¹

By Aboo Shoeb, Kanwal Raj, Randhir S. Kapil, and Satya P. Popli,* Central Drug Research Institute, Lucknow 226001, India

The structure and absolute configuration of *O*-methylalangiside (15) {11-(β -D-glycopyranosyloxy)-5,11,12,-12a,13,13a-hexahydro-2,3-dimethoxy-12-vinyl-6*H*-benzo[*a*]pyrano[3,4-*g*]quinolizin-8-one} has been established by chemical and spectroscopic studies and by correlation with ipecoside. Synthetic evidence is presented which indirectly confirms the 3-methoxy- (11) rather than the 2-methoxy-structure (12) for alangiside.

Alangium lamarckii Thw. (Alangiaceae) is a medicinal plant indigenous to India and is a rich source^{2,3} of alkaloids structurally related to the Ipecac bases, e.g. emetine (1). The isolation of dihydroprotoemetine (2),



ankorine (3),[†] and alangimarckine (4) added further interest to the chemical examination of this plant. The presence of the now familiar C₉₋₁₀ unit (shown as thickened bonds) derived presumably from loganin in all these structures led us to a search for the monoterpenoid glycosidic precursor^{1,4} predicted by biogenetic theory.

An extraction and isolation of the glycosidic fraction was accordingly undertaken on the lines described for the case of ipecoside⁵ (5). A brown amorphous powder was obtained which on purification by a combination of counter-current distribution and column chromatography gave a white amorphous powder shown to be pure by t.l.c. in two solvent systems and named alangiside.

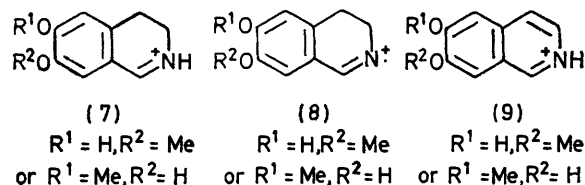
Alangiside could be isolated from the roots, leaves, or

[†] In view of recent work (C. Szántay, E. Szentirmay, and L. Szabo, *Tetrahedron Letters*, 1974, 3725), the structures of ankorine and alangimarckine may need to be revised with OH at position 8 instead of 11.

¹ Preliminary report, R. S. Kapil, A. Shoeb, S. P. Popli, A. R. Burnett, G. D. Knowles, and A. R. Battersby, *Chem. Comm.*, 1971, 904.

² A. R. Battersby, R. S. Kapil, D. S. Bhakuni, S. P. Popli, J. R. Merchant, and S. S. Saigar, *Tetrahedron Letters*, 1966, 4965.

fruit of *A. lamarckii* (best source unripe fruit). Its molecular formula, C₂₅H₃₁NO₁₀, was confirmed by mass spectrometric study of the compound itself and of four derivatives. Its u.v. spectrum [λ_{\max} (MeOH) 237 and 285 nm (log ϵ 4.27 and 3.68)], which underwent a bathochromic shift in alkali (λ_{\max} 247 and 303 nm) was similar to that of ipecoside.⁶ The i.r. spectrum indicated the presence of OH (ν_{\max} 3350br cm⁻¹) and only one carbonyl group (1650 cm⁻¹). The n.m.r. spectrum showed signals at τ 6.21 (OMe), 4.60 (3H, m, olefinic), 3.25 (2H, m, aromatic), 2.61 (1H, d, O=C-C=CH-O). The mass spectrum showed a weak *M*⁺ peak at *m/e* 505 and a stronger peak at *m/e* 343 (aglucone). The base peak was at *m/e* 274 (retro-Diels-Alder from *M*⁺) and there were other prominent peaks at *m/e* 272, 178, 177, and 176, the last three corresponding to the ions (7)–(9), respectively. The appearance of these



characteristic peaks in the mass spectrum of alangiside and its derivatives (see later) and the presence of dihydroprotoemetine, ankorine, and other benzoquinolizidine alkaloids in this plant pointed to the presence of a common benzoquinolizidine unit.

β -Glucosidase cleaved alangiside to yield D-glucose and the aglucone, C₁₉H₂₁NO₅. The mass spectrum of the aglucone showed ions at *m/e* 343 (*M*⁺) and 272 (retro-Diels-Alder), in agreement with the above findings. There were again characteristic peaks at *m/e* 178, 177, and 176.

The aglucone from *O*-methylalangiside, obtained in a similar manner, showed *M*⁺ 357, λ_{\max} 232 and 285 nm, and ν_{\max} 3300 and 1650 cm⁻¹. The mass spectrum showed the peaks corresponding to *m/e* 178 and 177 at *m/e* 192 and 191, respectively. The structural information obtained so far can be summarised in the partial formula (10).

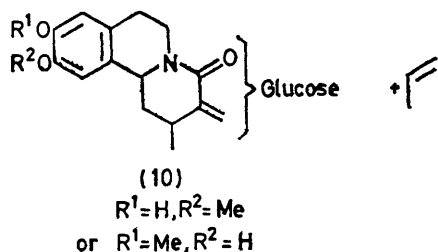
³ S. C. Pakrashi and B. Achari, *Experientia*, 1970, **26**, 933 and references therein.

⁴ S. P. Popli, Symposium on Chemistry of Natural Products, at Tashkent, Uzbek (U.S.S.R.), Sept. 16–20, 1968, Abstracts (supplement), p. 24.

⁵ P. Bellet, *Ann. pharm. franç.*, 1952, **10**, 81.

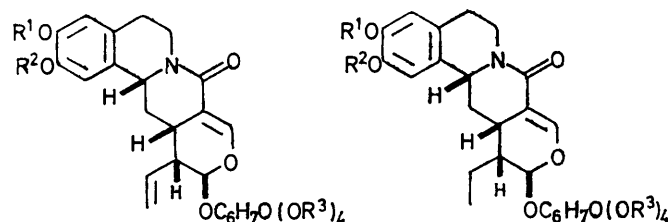
⁶ A. R. Battersby, B. Gregory, H. Spencer, J. C. Turner, M.-M. Janot, P. Potier, P. Francois, and J. Levisalles, *Chem. Comm.*, 1967, 219.

Treatment of alangiside with pyridine-acetic anhydride gave a penta-acetate (13), M^+ 715, ν_{\max} 1 750 cm^{-1} . The n.m.r. spectrum showed signals for the five



acetoxy-groups as well as two separate signals for the aromatic protons. In the mass spectrum, new peaks appeared at *m/e* 386, 368, and 331 (base peak). Catalytic hydrogenation of (13) furnished the dihydroalangiside penta-acetate (14).

Treatment of alangiside with diazomethane afforded *O*-methylalangiside (15) M^+ 519, acetylation of which gave the corresponding tetra-acetate (16) M^+ 687.



- (11) $R^2 = R^3 = \text{H}, R^1 = \text{Me}$
 (12) $R^1 = R^3 = \text{H}, R^2 = \text{Me}$
 (13) R^1 or $R^2 = R^3 = \text{Ac}, R^2$ or $R^1 = \text{Me}$
 (15) $R^1 = R^2 = \text{Me}, R^3 = \text{H}$
 (16) $R^1 = R^2 = \text{Me}, R^3 = \text{Ac}$
 (14) R^1 or $R^2 = R^3 = \text{Ac}, R^2$ or $R^1 = \text{Me}$

The foregoing data, in conjunction with biogenetic considerations, lead to structure (11) or (12) for alangiside.

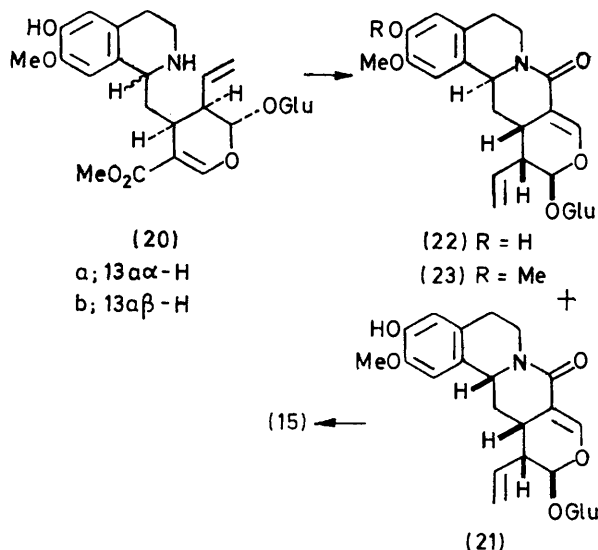
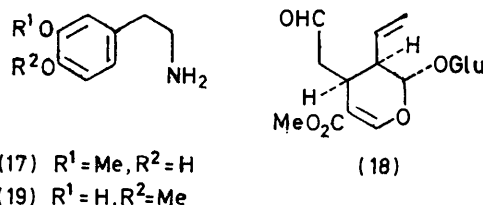
Correlation with deacetylpecoside (6) was therefore attempted (in the laboratories of Professor Battersby):¹ methylation of the lactam obtained from deacetylpecoside with diazomethane gave the ether (15), identical (spectroscopy and chromatography) with *O*-methylalangiside. Thus structure (15) is a complete representation of *O*-methylalangiside and either of the two structures (11) or (12) is admissible for alangiside.

To distinguish between these structures, a synthesis of alangiside was undertaken. At this stage, we were informed of an abundant source of the valuable intermediate, secologanin,⁷ from plants of the *Lonicera* species. The illustrated Scheme for the synthesis of the two isomers (11) and (12) was then devised.

Attempted condensation of 4-hydroxy-3-methoxyphenethylamine (17) hydrochloride with secologanin (18) under a variety of conditions was not successful. How-

⁷ Personal communications from C. R. Hutchinson and A. R. Battersby; cf. I. Souza and H. Mitsuhashi, *Tetrahedron Letters*, 1970, 191.

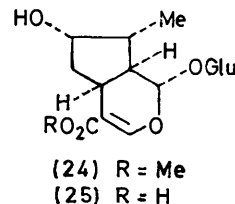
ever, 3-hydroxy-4-methoxyphenethylamine (19) condensed smoothly⁸ with secologanin to give the stereoisomers (20a and b). No attempts were made to obtain the individual epimers in pure form and the crude mixture was treated with a 15% solution of sodium carbonate



SCHEME

at room temperature for 30 min. The product consisted mainly of the β -epimer (21), which differed (optical rotation and n.m.r. spectrum) from alangiside and on treatment with diazomethane afforded a compound identical (spectra and t.l.c.) with naturally derived *O*-methylalangiside. This confirmed the stereochemistry of the synthetic product as well as the location of the phenolic hydroxy-group in alangiside (11).

If the reaction mixture was left for a longer time, besides the 13 β -isomer (21), the 13 α -isomer (22) could also be isolated; this was characterised as its *O*-methyl-ether (23).



In view of the importance of loganin (24) as an established precursor in the biosynthesis of the *Ipecacuanha*

⁸ A. R. Battersby, A. R. Burnett, and P. G. Parsons, *J. Chem. Soc. (C)*, 1969, 1187.

⁹ A. R. Battersby and B. Gregory, *Chem. Comm.*, 1968, 134; A. R. Battersby and R. J. Parry, *ibid.*, 1971, 901.

alkaloids⁹ as well as the bearing it has on the structure of alangiside, a search for this intermediate was also made in various parts of the plant collected in different seasons. Loganic acid (25) was isolated in 0.2% yield from the fresh unripe fruit of *A. lamarckii*.

EXPERIMENTAL

Isolation of Alangiside (II).—The pulverised fruits (3.0 kg) of *A. lamarckii* collected locally were percolated with ethanol (10 × 1.5 l), the extract was evaporated below 40° in *vacuo*, and the residue was diluted with water (500 ml). The solution was filtered, defatted with light petroleum ether, adjusted to pH 10 (Na₂CO₃), and extracted with ether–chloroform (4:1; 4 × 625 ml) to remove basic material. The aqueous layer was saturated with NaCl (60 g), brought to pH 4 (concentrated HCl), and extracted with chloroform–ethanol (4:1; 4 × 625 ml). The combined organic extract was washed with brine (50 ml), dried (Na₂SO₄), and concentrated to afford a residue (5.60 g), which was subjected to counter-current distribution (BuⁿOH–EtOAc–H₂O, 3:2:5; 10 ml fractions; 100 transfers). The contents of the tubes 55–69 were mixed on the basis of u.v. and n.m.r. data and afforded a cream-coloured powder which was further purified by column chromatography (silica gel; EtOAc–EtOH, 98:2) to furnish *alangiside* (2.25 g) as a white amorphous powder shrinking at 164° and melting at 187° (decomp.), $[\alpha]_D^{26} -105^\circ$ (*c* 1.0 in MeOH) ν_{\max} (KBr) 3 350br (OH), 1 650 (=N–C=O), 1 585, 1 510, 1 450, 1 440, 1 020, and 910 cm⁻¹, τ [(CD₃)₂SO: 60 MHz] 2.61 (1H, d, *J* 1.5 Hz, O–CH=), 3.25 (2H, s, ArH), 4.60 (3H, m, CH=CH₂), and 6.21 (3H, s, OMe), *m/e* 505 (*M*⁺), 343, 274 (base), 272, 178, 177, and 176 (Found: C, 54.25; H, 6.55; N, 2.3%; *M*⁺, 505.1925. C₂₆H₃₁NO₁₀·2.5H₂O requires C, 54.55; H, 6.55; N, 2.55%. C₂₅H₃₁NO₁₀ requires *M*, 505.1947).

Alangiside Penta-acetate (13).—A mixture of alangiside (500 mg), acetic anhydride (10.0 ml), and pyridine (5.0 ml) was kept overnight at room temperature. Solvents were removed under reduced pressure and the residue was diluted with water and extracted with benzene (3 × 10 ml). The extracts were combined, washed with water, dried (Na₂SO₄), and evaporated to afford the *penta-acetate* (13) (400 mg), which was further purified over a column of silica gel; ν_{\max} (CHCl₃) 1 750 (OAc) and 1 655 cm⁻¹ (=N–C=O), τ (CDCl₃) 2.54 (1H, d, *J* 2.5 Hz, O–CH=), 3.14 and 3.27 (1H each, both s, ArH), 5.75 (2H, m, CH₂-OAc), 6.18 (3H, s, OCH₃), 7.70 (3H, s, phenolic OAc), and 7.90, 7.97, 8.00, and 8.04 (3H each, all s, 4 OAc), *m/e* 715 (*M*⁺), 673, 386, 384, 368, 331 (base), 316, 220, 178, 177, 176, 170, 169, 127, and 109 (Found: C, 57.1; H, 5.95; N, 1.75%; *M*⁺, 715.2446. C₃₅H₄₁NO₁₅·H₂O requires C, 57.3; H, 5.85; N, 1.9%. C₃₆H₄₁NO₁₅ requires *M*, 715.2475).

Dihydroalangiside Penta-acetate (14).—A solution of the penta-acetate (13) (60 mg) in ethanol (25 ml) was stirred with Adams catalyst (30 mg) for 2 h in hydrogen. Filtration and concentration under reduced pressure afforded the *dihydro-derivative* (14) (58 mg), $[\alpha]_D^{26} -120^\circ$ (*c* 1.0 in MeOH), ν_{\max} (KBr) 1 750 and 1 655 cm⁻¹, τ (CDCl₃) 2.55 (1H, d, O–CH=), 3.08 and 3.22 (1H each, s, ArH), 5.74 (2H, m, CH₂-OAc), 6.15 (3H, s, OMe), 7.89, 7.95, 7.99, and 8.02 (1 phenolic OAc, 4 alcoholic OAc), and 9.01 (3H, t, CH₂Me), *m/e* 717 (*M*⁺), 675, 430, 371, 370, 369, 331, 298, 274, 272, 229, 178, 177, and 169 (Found: *M*⁺, 717.2568. C₃₅H₄₃NO₁₅ requires *M*, 717.2632).

O-Methylalangiside (15).—A solution of alangiside (150

mg) in the minimum quantity of methanol was treated with ethereal diazomethane [from *N*-nitrosomethylurea (350 mg)] and set aside overnight at room temperature. The solvent was removed and the residue (140 mg) was chromatographed over a column of silica gel to afford the *O-methyl derivative* (15) (70 mg), obtained as needles (from EtOAc), m.p. 236°, ν_{\max} (KBr) 3 375 and 1 655 cm⁻¹, τ (CDCl₃) 2.42 (1H, s, OCH=), 3.20 and 3.35 (1H each, s, ArH), 4.60 (3H, m, olefinic H), and 6.10 and 6.14 (each 3H, s, OMe), *m/e* 519 (*M*⁺), 505, 358, 357, 356, 340, 288 (base), 274, 272, 270, 258, 257, 256, 192, 191, 178, 177, 176, 149, and 145 (Found: C, 56.85; H, 6.85; N, 2.35%; *M*⁺, 519.2080. C₂₈H₃₃NO₁₀·1.5H₂O requires C, 57.15; H, 6.6; N, 2.55%. C₂₈H₃₃NO₁₀ requires *M*, 519.2104).

O-Methylalangiside Tetra-acetate (16).—A solution of *O*-methyl alangiside (15) (150 mg) in acetic anhydride (6 ml) and pyridine (3 ml) was left overnight at room temperature. The excess of reagent was removed under reduced pressure and the residue was diluted with water and extracted with benzene (3 × 15 ml). The extract was washed with water, dried (Na₂SO₄), and concentrated. The residue (120 mg) was chromatographed on a column of silica gel to furnish the *tetra-acetate* (16), ν_{\max} (KBr) 1 750 and 1 655 cm⁻¹, τ (CDCl₃) 2.49 (1H, d, OCH=), 3.34br (2H, s, ArH), 5.72 (2H, m, CH₂-OAc), 6.10 (6H, s, 2 OCH₃), and 7.89, 7.95, 7.99, and 8.00 (each 3H, s, 4 OAc), *m/e* 687 (*M*⁺), 400, 399, 356, 340, 331, 330, 288, 192, 191, 177, 169, 149, 145, 139, 127, 109, and 97 (Found: *M*⁺, 687.2502. C₃₄H₄₁NO₁₄ requires *M*, 687.2526).

Hydrolysis of Alangiside.—A suspension of alangiside (50 mg) and β-glucosidase (100 mg) in water (5 ml) and toluene (2 drops) was kept at room temperature for 3 days. It was then diluted with ethanol–chloroform (2:1:150 ml) and filtered. The filtrate was concentrated and the residue repeatedly extracted with chloroform (8 × 10 ml). Removal of the solvent afforded the *aglucone* (10 mg) as a white amorphous powder giving a single spot on t.l.c. (silica gel; EtOAc); ν_{\max} (KBr) 3 350br (OH), 1 655 cm⁻¹ (=N–C=O), *m/e* 343 (*M*⁺), 325, 314, 272, 243, 178, 177, and 176 (Found: *M*⁺, 343.1435. C₁₉H₂₁NO₅ requires *M*, 343.1419).

Hydrolysis of O-Methylalangiside.—By the procedure described above the corresponding *aglucone* was obtained as a light yellow amorphous powder, which when passed through a silica gel column yielded the pure product (t.l.c. on silica gel; EtOAc); ν_{\max} (KBr) 3 300br (OH) and 1 650 cm⁻¹ (=N–C=O), *m/e* 357 (*M*⁺), 342, 339, 328, 286, 272, 244, 242, 192, 191, 190, 177, and 176 (Found: *M*⁺, 357.1558. C₂₀H₂₃NO₅ requires *M*, 357.1576).

3-O-Demethyl-2-O-methylalangiside (21).—A mixture of 3-hydroxy-4-methoxyphenethylamine (19) hydrochloride (125 mg) and secologanin (18) (250 mg) in phosphate buffer (pH 5.2; 4.5 ml) was kept for 4 days at 37° under oxygen-free nitrogen. The reaction was monitored by t.l.c. The mixture was diluted with water (5 ml) and extracted with ethyl acetate (3 × 10 ml). The aqueous layer was further stirred at room temperature with aqueous 15% sodium carbonate (3 ml) for 30 min, then re-extracted with ethyl acetate, followed after 2.5 h by *n*-butanol (4 × 10 ml).

The ethyl acetate layer was concentrated and the residue (100 mg) chromatographed on a column of silica gel (5.0 g). Elution with 10% methanol–chloroform gave an amorphous powder (21), $[\alpha]_D^{25} -82.9^\circ$ (*c* 0.85 in MeOH), ν_{\max} (KBr) 3 350 and 1 650 cm⁻¹, τ [(CD₃)₂SO] 2.73 (1H, d, *J* 2.0 Hz, O–CH=), 3.20 (1H, s, ArH), 3.49 (1H, s, ArH), 4.60 (3H, m, CH=CH₂), and 6.25 (3H, s, OMe) (Found: *M*⁺, 505.1964. C₂₆H₃₁NO₁₀ requires *M*, 505.1947).

The n-butanol fraction on concentration gave a residue (200 mg) which was chromatographed on a column of silica gel (10.0 g). Elution with 10% methanol-chloroform afforded a powder (150 mg) [single spot on t.l.c. (silica gel; 20% MeOH in CHCl_3)]. Its n.m.r. spectrum indicated it to be a mixture of two components. The major one (65%) appeared to be (21) and the minor one (35%) was characterised as 3-O-demethyl-2-O-methylisoalangsaside (22), τ [(CD_3)₂-SO] 2.80 (1H, d, J 2.0 Hz, O-CH=), 3.20 (1H, s, ArH), 3.49 (1H, s, ArH), 4.60 (3H, m, CH=CH₂), and 6.25 (3H, s, OMe) (Found: M^+ , 505.1978).

O-Methylisoalangsaside (23).—A mixture of compounds (21) and (22) obtained from the foregoing n-butanol fraction was treated with an excess of diazomethane.

Column chromatography on silica gel (15.0 g) afforded a fraction largely consisting of compound (23), τ [(CD_3)₂SO] 2.74 (1H, d, J 2.0 Hz, OCH=), 3.10 (1H, s, ArH), 3.21 (1H, s, ArH), 4.60 (3H, m, CH=CH₂), and 6.21 (6H, s, 2 OMe) (Found: M^+ , 519.2120. $\text{C}_{26}\text{H}_{33}\text{NO}_{10}$ requires M , 519.2104). The later fractions from the column yielded O-methylalangsaside (15) (100 mg), m.p. 236°.

Isolation of Loganic Acid (25).—The n-butanol fraction (10 g) from the unripe green fruits (3.0 kg) of *A. lamarckii* was subjected to counter-current distribution (10 ml fractions; 100 transfers) in n-butanol-water (2 : 3). Tubes 26—36 afforded a fraction (850 mg) rich in (25). Part of the crude loganic acid (270 mg) was chromatographed on a column of silica gel (10 g); elution with ethyl acetate gave (25) (200 mg) as a pure white powder, λ_{max} (MeOH) 238 nm, λ_{max} (MeOH-0.1N-NaOH) 229 nm, ν_{max} (KBr) 3 350br (OH), weak bands in the region 2 700—2 500, 1 675br (C=O), and 1 625 cm^{-1} (C=C), τ (D_2O) 2.50br (1H, C:CH·O), 4.60 (1H, d, J 3 Hz, HC·C·OGlu), and 8.88 (3H, d, J 7 Hz, CHMe).

Loganin (24)—A solution of loganic acid (25) (200 mg) in methanol (2 ml) was treated with diazomethane [from *N*-nitrosomethylurea (250 mg)] and chromatographed on a column of silica gel (EtOAc) to yield pure loganin (120 mg), m.p. 222—223°, $[\alpha]_{\text{D}}^{25} - 83^\circ$ (c 1.0 in H_2O), identical with an authentic sample [mixed m.p. t.l.c. (silica gel; 10% EtOH- CHCl_3), and i.r. spectrum: ν_{max} (KBr) 3 500 (OH), 3 300 (OH), 1700 (CO_2Me), and 1 650 cm^{-1} (C=C)].

[4/2035 Received, 2nd October, 1974]